

" Understanding the role of gut microbiota in regulating the immune response and disease resistance in farmed fish: An numerical study using next-generation sequencing "

Reya Elanor^{1, *}

¹ The University of Auckland, New Zealand. *Corresponding author: Elanor.R@gmail.com

Abstract

The essential aim of this research study is to determine the role of gut microbiota in regulating immune response also that disease resistance in fish farmed. This research study represents that numerical analysis by using primary research data. For collecting the research data used, specific questions these questions fulfilled from new generation sequencing. This research study is also based on theoretical analysis for measuring the research using smart PLS software and running informative results, including the smart PLS Algorithm. The descriptive statistical analysis, the correlation coefficient, the R square, and the significant analysis also present the co-linearity statistical analysis between them. Overall results found that there are negative but significant links between the microbiota in regulating immune response and disease resistance in fish farmed.

Keywords: gut microbiota (GM), regulating immune response (RIR), disease resistance (DR), Fish Farmed (FF), Smart PLS Algorithm.

Citation: Young H. 2023. Evaluating the efficacy of a marine reserve in enhancing the population growth and size of endangered fish species. FishTaxa 27: 36-46.

Introduction

The Gut Microbiota

The gastrointestinal tract is a way designed by smooth muscle, submucosa, and mucosa, comprising a massive system of blood vessels and nerves. The mucosa controls for absorption and digestion of food from the nutrition and preservation of the body besides chemical and physical harm from luminal guts and source of immunity (Ghanbari et al., 2015). The small intestinal epithelium, the particular cell covers that methods the luminal aspect is doubled to form crypts and villi. This formation helps to raise the whole absorption surface of the intestine. Furthermore, in the mucosa, other kinds of cells exist, like enterocytes (the major cellular factor considers for around eighty percent), goblet, Paneth, and enter endocrine (including valid one percent of the epithelium). Enter endocrine cells, supplied during the gastrointestinal tract, method a large endocrine structure that manages: (a) gastrointestinal motility and secretion; (b) control of food ingestion; (c) postprandial glucose stages; and (d) through interaction with neurons innervating the gastrointestinal tract (these cells are dividing of the gut-brain axis). The mammalian intestine is occupied by a difficult group of microbes called microbiota, established by microorganisms, viruses, archaea (especially bacteriophages), fungi, and protozoans (Cénit et al., 2014). Shotgun metagenomic sequencing across unplanned sequencing of all genetic factors determined that the human gut influences a microbial microbiome that is dominated by phyla like Bacteroidetes, Actinobacteria, Proteobacteria Verrucomicrobia, and Firmicutes. Bacteroidetes and Firmicutes characterize ninety percent of the gut microbiota. It is vital to consider that the establishment of the human gut leads continues prenatally after birth, grasping the mature microbiota state between the ages of two and five years. Overall these data show that gut microbial establishment considerably affects the growth of the host immune system in initial life and influences disease and health in later life, as designated by the damage of immune function in germ-free mice (Parolini, 2019). An important alteration in the gut microbiota

arrangement accompanies the prevention of the introduction of solid foods and the mother by the improved establishment of butyrate manufacturers, comprising certain Clostridium species and Bacteroides. When well-known, the arrangement of the gut microbiota is comparatively constant through a mature lifetime in the lack of perturbations like the use of antibiotics, disease-associated dysbiosis, or long-term dietary changes (Yukgehnaish et al., 2020). Though centenarian microbiota presented reduced stages of Bacteroides, Enterobacteriaceae, and Bifid bacterium, Clostridium species stages were improved associated with younger adults. For these reasons, dietary-intervention investigations are necessary to examine whether altering the food arrangement of mature human beings can change their gut microbiota in a method that is helpful to their overall condition (Legrand et al., 2020).

The Gut Microbiota and the Immune System

A clear instance of the symbiotic results of the microbiome is the immune system, whose usual growth and actions are intensely affected by microbial metabolites that are created by: (a) Microbes from nutritional mechanisms; (b) biochemically and host adapted by gut bacteria; and (c) by gut microbes. In this situation, a difficult interaction between the intestinal epithelium, resident immune cells, and local microbiota has originated from developing, in which all contributors vigorously foster gastrointestinal homeostasis (Rimoldi et al., 2018). Segmented filamentous bacteria are top recognized for their capability to stimulate the differentiation of naive T cells to make antigen-specific Th-17 cells in the ileum of humans and mice. Th17 cells show a crucial part in the host protection in contradiction of extra-cellular pathogens, that are, extracellular bacteria and fungi and tissues homeostasis that raises epithelial blockade operations; however, they may cause auto-immunity (Kumar & Kocour, 2017). The methods occupied in stability among non-pathogenic and pathogenic Th-17 cells state are yet unidentified. But, medication of non-pathogenic Th-17 cells through IL23 alters it into a pathogenic phenotype, recommending that IL23 is a cytokine that impels the operational pheno-type of Th-17 cell.

Next-generation Sequence and fish gut microbiota

Numerous reports on mammals and humans have certainly associated gut microbiota groups with host nutrition, growth, and physiology. Fish gut microbiota is accountable for the production of intestinal enzymes, metabolism, and vitamins like short-chain (unstable) fatty acid, which signify the core energy resource for digestive epithelial cells. Additionally, the intestine harbor of fish is an extensive array of bacteria, mostly lactic acid bacteria, which may impede bacteria pathogens by concealing antimicrobial mixtures like acetic and lactic acid. Conversely, an unbalanced microbiota might destructively influence fish growth and nutrition and direct a modification of the gut immune system, participating so to the growth of diseases (Olsvik et al., 2013). Thus, an improved perception of microbe/gut connections and gut microbiota range in fishes might be extremely related to aqua-culture application. Fish microbiota conventionally deliberate by consequent detection and culture approaches founded on pheno-typic and biochemical properties of bacteria. But, techniques that depend on culture offer a partial image of intestine microbiota as just a less ratio, downwards to around one percent of bacteria from the fish intestine may be grown. So, in the recent era, some cultureindependent molecular methods have been established and functional to investigate the fish gut microbiota (Houston, 2017). The most influential method to analyze the configuration of multifaceted intestine microbial groups is characterized by a technology called Next-generation Sequencing. Metagenomic describing through a higher amount sequence of cpn-60 gene or 16S-rRNA was functional in several recent reports to analyze the influence of nutritional plant components on the fish gut microbiota configuration.

In the field of fisheries, next-generation sequencing approaches are utilized to study transcriptome-wide and genome-wide and regulate biological procedures, recognition of new signs for populace assembly, phylogenetics, traceability, genetic charting and ecotoxicological uses, and study of the connotation of loci

with characters influenced by choice. The usage of next-generation sequencing has allowed investigators to get a greater extent of resolution than utilizing conventional genetic signs, so revealing the evidence is not achievable earlier. In spite of the favorable consequences got utilizing next-generation sequencing approaches, it has been used in fisheries reports on partial no. of fish class (Franco et al., 2021).

Research objective

The main aim of this research is to study the biological perceptions obtained from next-generation sequencing approaches for a diversity of fish classes in accordance to achieve a complete perspective of this method in the near future.

Literature review:

Studies revealed that gut microbiota plays a significant role in host immune system regulation. by the modulation of gut microbiota, fish health can be controlled. The understanding of the interaction of hostmicrobiota and the after-effects of diseases are treated using probiotics. Probiotics provide resilience against disease as well as increase the chances of speedy recovery (Rasmussen et al., 2022).studies explained that the expression of certain immune-related genes increases in the intestine through the dietary supplements of Yu-Ping-Feng (YPS). YPS has no significant effect on the growth rate of shrimps, and thus, it is suitable for aquaculture-based framed shrimps(Su et al., 2020).studies show that the study of microbiomes involves dealing with microorganisms related to the gut of human beings or animals. The microbes provide defense and strengthen the immune system in response to the many diseases condition. For studying and understanding the role of the microbiome at the species level, various next-generation sequencing technology is used for research purposes. these next generation-based technologies include an Amplicon as well as a Shotgun. Both these technology works on the principle of bioinformatics for providing information about the microbe present at the specie level. studies explain that the microbiome present in the fish gut plays a role in food digestion and provides nutrition and disease vulnerability features to the host. Also, the development and innovation of bioproducts are possible due to microbial-based communities (Sahu et al., 2008). Studies using the Gnotobiotic Zebrafish Model predicted that it is identified that gut microbiota plays a critical role in the development process of the gut as well as in immune-based responses(Ou et al., 2021).studies claim that using the lactic acid bacteria for increasing the efficacy of turbot Scophthalmus Maximus plays its role in intestinal microbiota and affect the expression of immune-associated genes(Guo et al., 2020).studies show that the growth and immunity associated factors of farmed fish are promoted using the probiotics. probiotics work by mediating the mechanism of host-microbiota interactions. for the understanding of the molecular interaction of host-microbiome, the Rainbow Trout Model is used in research studies. The model investigates the interaction using two probiotic bacteria (Zhao et al., 2023).studies explained that for providing information about the gut profile and its relation with metabolites when JCM5805-supplement is applied then the technique of gas chromatography is used in the study work. the result of the research work showed that JCM5805 affects the host growth as well as an immunity when applied to studying the relationship between metabolites and gut microbiota(Xia et al., 2020).studies claim that microbial interaction with the intestine helps in maintaining host health. The studies r provides an understanding of the intestinal bacteria in regulating the activity of shrimp intestines using probiotics function(Du et al., 2019).studies revealed that fish mucosal surface provides a barrier that works by regulating the diversity of microbial assemblages. Regulating the microbial assemblages improves the host's health and fitness. for managing health conditions, farmed species' microbial system is regulated through several stressors(Li et al., 2022).studies explain the role of short-chain fatty acids in improving the modulation of gut microbiota. these SCFAs cause immune response enhancement and increase the host antioxidant capacity(Lin et al., 2020).studies show the change in gut health due to the effects of dietary fibers and fish fiber consumption on the immune .dietary fibers alter the diversity associated with

bacteria's thereby modulating the community structure. The use of moderate fiber supplements is beneficial for promoting fish intestinal health. The antioxidants and other anti-inflammatory effects associated with moderate fibers enhance the intestinal health-promoting features in fish(Lee et al., 2021).studies of scholars claim that the metabolism of host fish is affected due to the gut microbiome of fish. When the utilization of antimicrobial-resistant bacteria is combined with gut microbiomes, it promotes the positive use of these microbiomes as probiotics. The effective alternative to antibiotics is probiotics that act as a potential and beneficial microbe. form various metagenomic studies, it is revealed that the bioinformatic technology-based system provides information about the effect of the gut-associated microbiome on fish physiology(Meng et al., 2021).studies predicted that immune homeostasis is maintained through gastrointestinal immune responses. theses hemostatic regulation maintains the host-microbiome interactions by triggering innate as well as host adaptive immunity. These interactions then provide a defense mechanism to the host immune system. The administration of oral administrate immunoprophylactic helps in improving the health of fish.in aquaculture industries, these immunoprophylactic provide disease management feature(Liu et al., 2022).various studies explained that dietary fibers affect intestinal physiology and the expression of gene related to the immune system .the result of the studies claim that supplements along with Clostridium Butyricum when given to common carp in proper doses, improves its intestinal immunity. the main reason behind intestinal immunity improvement in carp is that these doses regulate the intestinal microbiota defense functioning, thereby improving the intestinal response (López Nadal et al., 2020).studies have regarded Lipopenaeus Vannamei as the shrimp species with the most productivity around the globe. Due to adverse environmental factors, the shrimp population is decreasing because the outbreak affects the immune system of shrimps and causes immune-related disease in them .for improving the function of shrimps, Lactobacillus Pentosus combined with Arthrospira platensis is used as a substitute for antibiotics(Ouyang et al., 2023).studies provide evidence about the functioning of zebrafish gut and molecular pathways associated with zebrafish gut. The zebrafish model is used for studying the feeding effects of fish species and for studying the immune interaction linked with hostmicrobiome(Yu et al., 2023).scholars predicted that in Mandarin fishes, the saline and alkali water influence their health and immune-related functions. Also, studies Explain the effect of the relation of intestinal flora with complex host environment and its influence on disease resistance phenomenon(Holt et al., 2021).studies revealed that Stress Worry-Free Concentrate Probiotics(SWFC) play a crucial role in improving additives that are essential for fish health. The main application of SWFC is that it regulates the inflammatory responses associated with the immune system of the host. the use of SWFC provides numerous positive effects on mucus as well as on gut functionality. also, the postbiotic effect of SWFC, when fed with a high-fat diet, improves the microbial structure and function in common carp (Wang et al., 2023).studies explained the microbiome's involvement in regulating shrimp health. The functioning of the shrimp microbe gets altered with shrimp pathogens' introduction in the host. The role of microbiome supplements in the production of penaeid shrimp holds significant value(Luna et al., 2022).studies highlighted that Tilapia immunity and health are improved using the improved strategies scholars predicted that the approach of biotechnology in microbiome research holds great value for improving intestinal immune-associated diseases.

Disease:

Bacterial populations in the digestive system, according to a study, contribute to the development of gut diseases such as inflammatory bowel disease (IBD) such as Crohn's disease, and ulcerative colitis. Low microbial diversity in the gut has also been related to obesity and type 2 diabetes. According to a credible source, the status of the gut microbiota is linked to metabolic syndrome. Prebiotics, probiotics, and other dietary supplements can help to change the diet and reduce these risk factors. Antibiotic usage that disrupts the microbiota can lead to disease, including antibiotic-resistant diseases. Another important role of the microbiota

is the intestinal overgrowth of externally introduced populations that would otherwise cause disease; the "good" bacteria compete with the "negative," and some even release anti-inflammatory substances. Animals' digestive systems are home to a variety of microorganisms called gut microbiota, gut microbiome, or gut flora. These species include bacteria, archaea, fungi, and viruses. The collective genomes of the gut bacteria make up the gastrointestinal metagenome. The stomach contains the majority of the human microbiome. The gut microbiota performs a variety of tasks, such as colonization, pathogen resistance, maintaining the intestinal epithelium, metabolizing food and medication, regulating immune system activity, and even controlling behavior via the gut-brain axis.

The gut microbiota's microbial composition varies in various regions of the digestive tract. With between 300 to 1000 different species present, the colon has the most microorganisms of any habitat on Earth. Bacteria, which comprise 30 or 40 species and represent 99% of gut bacteria, are now the most studied and prevalent component. Up to 60% of the dry mass of feces might be made up of bacteria. Large quantities of aerobic bacteria may be detected in the cecum, despite the fact that more than 99% of the bacteria in the gut are anaerobes. It is estimated that the human gut microbiota has a hundred times greater number of genes than the human genome.

The composition of the gut microbiome is influenced by the geographic origin of populations. Prevotella trade-off variations, urease gene representation, and deviations in the expression of genes recording glutamate, which protease/degradation or additional enzymes involved in amino acid degradation or vitamin biosynthesis reveal significant differences between populations from the United States, Malawi, and Amerindian origin. In contrast to the US population, Malawian and Amerindian populations have high representations of glutamate synthase-encoding enzymes as well as an excess of -amylase in their microbiomes. These populations also have high representations of enzymes involved in the vitamin and lipoic acid biosynthesis. The diet is likely the primary predictor of the gut bacterial makeup because the US population has a diet higher in lipids than Amerindian or Malawian populations, whose diets are high in maize.

More research has revealed that youngsters in rural Africa and Europe have very different microbiota compositions. youngsters from the small rural community of Boulpon in Burkina Faso and youngsters from Florence had their fecal microorganisms compared. A typical villager's child has a diet that is mainly deficient in fats and animal proteins and abundant in carbohydrates and plant proteins. While Firmicutes predominated in the feces of European children and exhibited a significant decline in biodiversity, Bacteroidetes predominated in the feces of the Boulpon children. Increased biodiversity and a diverse gut microbiome makeup in African cultures may help digest plant polysaccharides that are often indigestible and may also lead to a decrease in the prevalence of non-infectious colonic diseases.

On a smaller scale, it has been demonstrated that the composition of a person's microbiome is strongly influenced by the number of similar environmental exposures that members of the same family experience. This effect is routinely seen in populations from various cultural backgrounds and has no genetic basis.

Humans acquire a gut flora similar to adults between one to two years after birth. Along with the development of the gut flora, the intestinal epithelium and the intestinal mucosal barrier that it secretes grow in a way that is somewhat tolerant of and even supportive of commensalism bacteria while simultaneously functioning as a barrier against pathogenic ones. To create an outside mucosal layer where "friendly" bacteria may anchor and feed and an internal mucosal layer where even these organisms cannot penetrate, the mucosa-producing goblet cells in particular, increase, and the mucosa layer develops. Additionally, the development of gut-associated lymphoid tissue (GALT), a part of the intestinal epithelium that recognizes and reacts to pathogens, corresponds with the formation of the gut flora. Although it is tolerant to some species of gut flora, the GALT that develops is resistant to other microorganisms. GALT typically develops a tolerance to the food the

newborn is exposed to, as well as to the meal's digestive byproducts and the metabolites (molecules created during metabolism) produced by the gut flora.

In order to protect itself, the human immune system produces cytokines that may either stimulate inflammation or dampen it in order to preserve homeostasis and promote healing following shock or damage. While certain segmented filamentous bacteria appear to promote the generation of inflammatory cytokines, several Clostridia species and Bacteroides fragilis appear to promote an anti-inflammatory response. This shows that specific bacterial species found in gut flora might affect the immune system's ability to produce cytokines. The immune system's ability to produce antibodies can also be controlled by the flora in the gut. This modulation has the effect of changing the class of B cells from B to IgA. In most situations, T helper cells must activate B cells in order to promote class flipping; however, in another pathway, gut flora causes intestinal epithelial cells to activate NF-kB, which causes the release of additional signaling molecules. These signaling molecules engage B cells in an interaction that results in IgA class switching. In mucosal habitats, such as the gut, IgA is a crucial form of antibody. It has been demonstrated that IgA can assist the gut flora in becoming more diverse and aids in the elimination of bacteria that trigger inflammatory reactions. In the end, IgA keeps the host and gut flora in a healthy environment. These cytokines and antibodies can impact the lungs and other organs in addition to the stomach.

Numerical analysis:

Name	No.	Туре	Missings	Mean	Median	Observed min	Observed max	Standard deviation	Excess kurtosis	Skewness	Cramér- von Mises p value
GM	0	MET	0	1.586	2.000	1.000	3.000	0.620	-0.576	0.572	0.000
RIR	1	MET	0	1.727	2.000	1.000	4.000	0.827	0.358	0.987	0.000
DR	2	MET	0	1.535	1.000	1.000	4.000	0.743	0.337	1.153	0.000

Table-1

The above result represents that descriptive statistic analysis. The result describes overall mean values, standard deviation, skewness values, and probability values. The result represents that the mean value of gut microbiota is 1.586. Its standard deviation value is 0.620, showing that 62% deviate from mean values. The result also represents that the skewness value is 0.572, showing that 57% skewness rates the RIR represent that the mean value is 1.727. Its standard deviation rate is 0.82, showing that 82% deviate from mean values. The overall probability value shows that 100% significant level between them. The result also describes that DR values its mean rate is 1.535 the standard deviation rate is 74% deviating from the mean. According to the result, its skewness value presents that 1.153 respectively.

Correlation coefficient:

	GM	RIR	DR	
DR	-0.023	-0.042	1.000	
GM	1.000	0.000	0.000	
RIR	-0.023	1.000	0.000	

Table-2

The above result represents that the correlation coefficient result describes the negative rate between GM and DR; its value is -0.023. Itsit also presents a negative rate with RIR; its value is -0.042, respectively, showing the negative values between them. the GM shows that 100% significant effect with RIR and 100% interrelation with each other.

Path coefficient analysis:

variables	Path coefficients	Alpha 1% power 80%	Alpha 5% power 80%	Alpha 1% power 90%	Alpha 5% power 90%
Regulating the immune response->	0.042	5723.000	3526.000	7423.000	4884.000

disease resistance					
Gut microbiota-> Regulating the immune	0.0231	18479.00	11384.00	23967.000	15768.000
response					

Table-3

The above result represents that path coefficient analysis result describes that path coefficient values, the alpha 1% and 80% power values, between regulating the immune response and disease resistance its rates are 0.042, 5723.000, 3526.000, 7423.000, and 4884.000, respectively. The gut microbiota-> and regulating the immune response present that path coefficient value is 0.0231, its alpha of 1%, power 80% rate is 18479.00, its alpha of 5% power 80% rate is 11384.00, respectively shows that positive rates between them.

Model selection criteria:

VARIABLES	BIC (bayesian information criterion)
Regulating the immune response	8.131
Disease resistance	8.011

Table-4

The above result describes the model selection criteria. Its BIC values are 8.131 and 8.011 for regulating the immune response and disease resistance.

Model fitness analysis:

SATURATED MODEL	ESTIMATE MODEL
0.010	0.010
0.000	0.001
0.004	0.000
0.654	0.543
1.000	0.799
	0.010 0.000 0.004 0.654

Table-5

The above result represents that the model fitness analysis result describes the saturated model and estimated model. The SRMR value is 0.010; its estimate model is 0.010, respectively. The chi-square value of the saturated model is 0.654, and the estimated model value is 0.543, showing that 65% and 54% model fitness analysis between them. The result also presents that the NFI value is 1.000 and the estimated model value is 0.799, showing 79% fitness rates between them.

Co-linearity statistic analysis:

Variables	VIF
DR GM	1.000
GM	1.000
RIR	1.000

Table-6

The above result represents that co-linearity statistic analysis and its VIF values are 1.000, respectively, showing that there are 100% linearity effects between them.

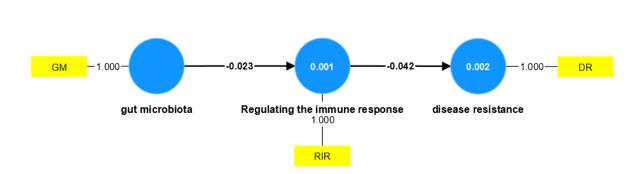
R-squares:

R-square	R-square adjusted
0.654	0.410
0.657	0.542
	0.654

Table-7

The above result describes that R square values also that adjusted R square values the rate of regulating the immune response its r square value is 0.654, and its r square adjusted value is 0.410 respectively. The result shows that 65% and 41% of research is reliable for analysis related to the gut microbiota in regulating the immune response. Also, that disease resistance, according to the result, its R square value is 65%, and its adjusted R square value is 54%, respectively.

Smart PLS Algorithm Model:



The above algorithm model represents that the smart PLS Algorithm is present in that gut microbiota and regulating the immune response show a negative link with each other. Disease resistance and regulating the immune response also show a negative relationship between them. The rate is -0.042 respectively.

Significant analysis:

	Original sample (0)	Sample mean (m)	Standard deviation (STDVEV)	T statistics (10/STDEVI)	P values
DR<- disease resistance	1.000	1.9032	1.8782	1.9021	0.000
GM<-gut microbiota	1.000	1.3445	1.8921	1.9999	0.000
RIR<-Regulating the immune response	1.000	1.2345	1.9021	1.98821	0.000

Table-8

The above result describes that significant analysis results represent the original sample value, simple mean values, standard deviation values, and also that T statistic values, and probability values. The result represents that the first matrix is DR<-disease resistance. Its original sample value is 1.000; its mean sample value is 1.9032. Its standard deviation value is 1.8782. according to the result, its t statistic value is 1.9021, and its probability value is 0.000, showing that positive and 100% significant analysis between the DR<-disease resistance. The second matrix is GM<-gut microbiota. According to the result, its original sample value is 1.000 the sample mean value is 1.234. the standard deviation represents 1.9021; its t-statistic value is 1.98. also, the significant value is 0.000, showing that a 100% significant level between them related to the gut microbiota in regulating the immune response and disease resistance.

The immune system is a collection of cellular and humoral elements that work in response to endogenous or exogenous stimuli to protect the body from outside invaders like germs, poisons, or cancerous cells. Although it is now recognized that these two immune systems collaborate to eliminate intruders or activate defense mechanisms, the fish immune system is classified into two categories: innate and adaptive (memory), both of which are further separated into humoral factors (soluble substances) and cell-mediated defense. The innate system, which works as the body's initial line of defense and reacts more quickly than the specialized system, consists of all elements that were there in the body before the pathologic agent appeared. Along with the complement system, antimicrobial enzymes, interleukins, interferon, and organic defense cells, including granulocytes, monocytes, macrophages, and natural killer cells, the skin functions as a physical barrier. The majority of the time, the balance between host defense and immunological tolerance is maintained by the gut bacteria. There is evidence of immune system changes to dysbiosis of the gut microbiota. This review focuses on the mutual interaction between the innate immune compartment and the gut microbiota, with a particular focus on the innate lymphoid tissue, innate lymphoid cells, and phagocytes. The review offers a potential clinical explanation for how the "gut microbiota—innate immunity" axis may play a role in the etiology of

autoimmune conditions such as rheumatoid arthritis, spondyloarthritis, and systemic lupus erythematosus.

The complex interactions between the local innate immune system and the gut bacteria define the microenvironment in the gut. GALTs' structural development and the priming of different immune cells for effective immunological activity depend on commensal bacteria. The procedures include a discussion of epigenetic alteration employing metabolic byproducts like SCFAs, antigen exposure and presentation, and signaling pathways based on the identification of PRR-PAMPs. Commensal flora and a strong immune system, according to current views, are necessary for self-tolerance, but dysbiosis may alter the local immune system and compromise tolerance. ILCs and phagocytes, including macrophages and dendritic cells, are crucial for regulating self-reacting immune cells, according to studies.

On the other hand, there is sufficient evidence to draw the conclusion that gut microbiota dysbiosis is a comorbidity of autoimmune diseases such as systemic lupus erythematosus, rheumatoid arthritis, and spondylarthritis. Dietary variables, genetic predisposition, immune system issues, or combined may influence the gut microbiota. Dysbiosis in the preclinical stage, however, may start or aggravate immunological diseases, according to other studies. Although the "microbiota innate immunity disease" axis has not yet been fully established, it may represent a significant pathogenic mechanism in a number of diseases. Combining traditional therapies for autoimmune diseases with early management to reverse the dysbiosis state by fecal transplantation or probiotic supplementation may be a therapeutic approach.

Conclusion:

Contradictory findings have been reached from a small number of research, however, There are still questions about the viability and effectiveness of such techniques, but several ongoing large cohort clinical trials may rapidly offer additional resistant. Going one step further, understanding the functions of the immune system and gut microbiota may lead to a significant advancement in the treatment of autoimmune diseases. The gut microbiota is the collection of microorganisms that lives in a person's gastrointestinal tract. This includes a wide variety of bacteria, fungi, viruses, and other organisms. For the human body to produce energy, digest food, and mount an immune defense, the digestive system and gut bacteria must cooperate. Intricate networks of microbiotas may be found in the epidermis, colon, vaginal, and oral cavities, among other places in the body. The term "human microbiome" refers to the total number of microorganisms in the body. In conclusion, immunostimulants may not be effective against all infections, but they help lessen the losses brought on by different diseases in aqua-culture. The timing, doses, manner of management and health state of the animal must all be considered for the use of immunostimulants to be effective. In aqua-culture, immunostimulants are drugs that prevent disease. They increased the bactericidal activities of phagocytic cells and improved their functionality. Additionally, they promote and enhance fish lysozyme and antibody responses. Injecting immunostimulants into fish is the most efficient means of doing consequently. It has also been shown that oral and immersion treatments work, although their effectiveness wanes with time.

Reference

- Cénit, M., Matzaraki, V., Tigchelaar, E., & Zhernakova, A. (2014). Rapidly expanding knowledge on the role of the gut microbiome in health and disease. *Biochimica et Biophysica Acta (BBA)-Molecular Basis of Disease*, 1842(10), 1981-1992.
- Du, Y., Zhou, S., Liu, M., Wang, B., Jiang, K., Fang, H., & Wang, L. (2019). Understanding the roles of surface proteins in regulation of Lactobacillus pentosus HC-2 to immune response and bacterial diversity in midgut of Litopenaeus vannamei. Fish & shellfish immunology, 86, 1194-1206.
- Franco, C. M., Ambrosio, R. L., Cepeda, A., & Anastasio, A. (2021). Fish intended for human consumption: From DNA barcoding to a next-generation sequencing (NGS)-based approach. *Current Opinion in Food Science*, 42, 86-92.
- Ghanbari, M., Kneifel, W., & Domig, K. J. (2015). A new view of the fish gut microbiome: advances from next-generation sequencing. *Aquaculture*, 448, 464-475.

- Guo, G., Li, C., Xia, B., Jiang, S., Zhou, S., Men, X., & Ren, Y. (2020). The efficacy of lactic acid bacteria usage in turbot Scophthalmus maximus on intestinal microbiota and expression of the immune related genes. *Fish & shellfish immunology*, 100, 90-97.
- Holt, C. C., Bass, D., Stentiford, G. D., & van der Giezen, M. (2021). Understanding the role of the shrimp gut microbiome in health and disease. *Journal of invertebrate pathology*, *186*, 107387.
- Houston, R. D. (2017). Future directions in breeding for disease resistance in aquaculture species. *Revista Brasileira de Zootecnia*, 46, 545-551.
- Kumar, G., & Kocour, M. (2017). Applications of next-generation sequencing in fisheries research: A review. Fisheries Research, 186, 11-22.
- Lee, P.-T., Yamamoto, F. Y., Low, C.-F., Loh, J.-Y., & Chong, C.-M. (2021). Gut immune system and the implications of oral-administered immunoprophylaxis in finfish aquaculture. *Frontiers in immunology*, *12*, 773193.
- Legrand, T. P., Wynne, J. W., Weyrich, L. S., & Oxley, A. P. (2020). A microbial sea of possibilities: current knowledge and prospects for an improved understanding of the fish microbiome. *Reviews in Aquaculture*, *12*(2), 1101-1134.
- Li, S., Heng, X., Guo, L., Lessing, D. J., & Chu, W. (2022). SCFAs improve disease resistance via modulate gut microbiota, enhance immune response and increase antioxidative capacity in the host. *Fish & shellfish immunology*, *120*, 560-568.
- Lin, S.-M., Zhou, X.-M., Zhou, Y.-L., Kuang, W.-M., Chen, Y.-J., Luo, L., & Dai, F.-Y. (2020). Intestinal morphology, immunity and microbiota response to dietary fibers in largemouth bass, Micropterus salmoide. *Fish & shellfish immunology*, 103, 135-142.
- Liu, L., Cai, X., Ai, Y., Li, J., Long, H., Ren, W., Huang, A., Zhang, X., & Xie, Z.-y. (2022). Effects of Lactobacillus pentosus combined with Arthrospira platensis on the growth performance, immune response, and intestinal microbiota of Litopenaeus vannamei. *Fish & shellfish immunology*, *120*, 345-352.
- López Nadal, A., Ikeda-Ohtsubo, W., Sipkema, D., Peggs, D., McGurk, C., Forlenza, M., Wiegertjes, G. F., & Brugman, S. (2020). Feed, microbiota, and gut immunity: using the zebrafish model to understand fish health. *Frontiers in immunology*, 11, 114.
- Luna, G. M., Quero, G. M., Kokou, F., & Kormas, K. (2022). Time to integrate biotechnological approaches into fish gut microbiome research. *Current Opinion in Biotechnology*, 73, 121-127.
- Meng, X., Wu, S., Hu, W., Zhu, Z., Yang, G., Zhang, Y., Qin, C., Yang, L., & Nie, G. (2021). Clostridium butyricum improves immune responses and remodels the intestinal microbiota of common carp (Cyprinus carpio L.). *Aquaculture*, 530, 735753.
- Olsvik, P. A., Vikeså, V., Lie, K. K., & Hevrøy, E. M. (2013). Transcriptional responses to temperature and low oxygen stress in Atlantic salmon studied with next-generation sequencing technology. *BMC genomics*, 14, 1-21.
- Ou, W., Yu, G., Zhang, Y., & Mai, K. (2021). Recent progress in the understanding of the gut microbiota of marine fishes. *Marine Life Science & Technology*, *3*, 434-448.
- Ouyang, H., Deng, N., Xu, J., Huang, J., Han, C., Liu, D., Liu, S., Yan, B., Han, L., & Li, S. (2023). Effects of hyperosmotic stress on the intestinal microbiota, transcriptome, and immune function of mandarin fish (Siniperca chuatsi). *Aquaculture*, *563*, 738901.
- Parolini, C. (2019). Effects of fish n-3 PUFAs on intestinal microbiota and immune system. Marine drugs, 17(6), 374.
- Rasmussen, J. A., Villumsen, K. R., von Gersdorff Jørgensen, L., Forberg, T., Zuo, S., Kania, P. W., Buchmann, K., Kristiansen, K., Bojesen, A. M., & Limborg, M. T. (2022). Integrative analyses of probiotics, pathogenic infections and host immune response highlight the importance of gut microbiota in understanding disease recovery in rainbow trout (Oncorhynchus mykiss). *Journal of Applied Microbiology*, 132(4), 3201-3216.
- Rimoldi, S., Terova, G., Ascione, C., Giannico, R., & Brambilla, F. (2018). Next generation sequencing for gut microbiome characterization in rainbow trout (Oncorhynchus mykiss) fed animal by-product meals as an alternative to fishmeal protein sources. *PLoS One*, *13*(3), e0193652.
- Sahu, M. K., Swarnakumar, N., Sivakumar, K., Thangaradjou, T., & Kannan, L. (2008). Probiotics in aquaculture: importance and future perspectives. *Indian journal of microbiology*, *48*, 299-308.
- Su, C., Fan, D., Pan, L., Lu, Y., Wang, Y., & Zhang, M. (2020). Effects of Yu-Ping-Feng polysaccharides (YPS) on the immune response, intestinal microbiota, disease resistance and growth performance of Litopenaeus vannamei. Fish & shellfish immunology, 105, 104-116.
- Wang, B., Thompson, K. D., Wangkahart, E., Yamkasem, J., Bondad-Reantaso, M. G., Tattiyapong, P., Jian, J., & Surachetpong, W. (2023). Strategies to enhance tilapia immunity to improve their health in aquaculture. *Reviews in Aquaculture*, 15, 41-56.
- Xia, Y., Yu, E., Lu, M., & Xie, J. (2020). Effects of probiotic supplementation on gut microbiota as well as metabolite

profiles within Nile tilapia, Oreochromis niloticus. Aquaculture, 527, 735428.

- Yu, Z., Hao, Q., Liu, S.-B., Zhang, Q.-S., Chen, X.-Y., Li, S.-H., Ran, C., Yang, Y.-L., Teame, T., & Zhang, Z. (2023). The positive effects of postbiotic (SWF concentration®) supplemented diet on skin mucus, liver, gut health, the structure and function of gut microbiota of common carp (Cyprinus carpio) fed with high-fat diet. *Fish & shellfish immunology*, 135, 108681.
- Yukgehnaish, K., Kumar, P., Sivachandran, P., Marimuthu, K., Arshad, A., Paray, B. A., & Arockiaraj, J. (2020). Gut microbiota metagenomics in aquaculture: Factors influencing gut microbiome and its physiological role in fish. *Reviews in Aquaculture*, 12(3), 1903-1927.
- Zhao, C., Men, X., Dang, Y., Zhou, Y., & Ren, Y. (2023). Probiotics Mediate Intestinal Microbiome and Microbiota-Derived Metabolites Regulating the Growth and Immunity of Rainbow Trout (Oncorhynchus mykiss). *Microbiology* Spectrum, 11(2), e03980-03922.